

This Page Is Inserted by IFW Operations  
and is not a part of the Official Record

## BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,  
please do not report the images to the  
Image Problems Mailbox.**

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION  
International Bureau

## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6:  G01N		A2	(11) International Publication Number: <b>WO 97/26523</b>
			(43) International Publication Date: <b>24 July 1997 (24.07.97)</b>
<p>(21) International Application Number: <b>PCT/US97/00881</b></p> <p>(22) International Filing Date: <b>17 January 1997 (17.01.97)</b></p> <p>(30) Priority Data: 08/588,526 18 January 1996 (18.01.96) US</p> <p>(71) Applicant: PROGENITOR, INC. [US/US]; 1507 Chambers Road, Columbus, OH 43212 (US).</p> <p>(72) Inventors: SNODGRASS, H., Ralph; 650 Retreat Lane, Powell, OH 43065 (US). CIOFFI, Joseph; 1180 Bayboro Drive, New Albany, OH 43054 (US). ZUPANCIC, Thomas, J.; 501 Park Boulevard, Worthington, OH 43085 (US). SHAFFER, Alan, W.; 256 Lakeview Drive, Lancaster, OH 43130 (US).</p> <p>(74) Agents: POISSANT, Brian, M. et al.; Pennie &amp; Edmonds, 1155 Avenue of the Americas, New York, NY 10036 (US).</p>			<p>(81) Designated States: AL, AM, AU, AZ, BA, BB, BG, BR, BY, CA, CN, CU, CZ, EE, GE, HU, IL, IS, JP, KG, KP, KR, KZ, LC, LK, LR, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, TJ, TM, TR, TT, UA, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p><b>Published</b> <i>Without international search report and to be republished upon receipt of that report.</i></p>
<p>(54) Title: DETECTION OF A LEPTIN RECEPTOR VARIANT AND METHODS FOR REGULATING OBESITY</p> <p>(57) Abstract</p> <p>The present invention relates to a variant form of the receptor for the <i>obese</i> gene product. In particular, the invention relates to methods of detecting this receptor variant in cells and tissues of obese individuals. In addition, it relates to methods of inhibiting or down-regulating expression of this variant in cells to augment their responsiveness to weight regulation by leptin as well as methods of using compounds to directly activate signal transduction pathways associated with this ligand-receptor system.</p>			

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AM	Armenia	GB	United Kingdom	MW	Malawi
AT	Austria	GE	Georgia	MX	Mexico
AU	Australia	GN	Guinea	NE	Niger
BB	Barbados	GR	Greece	NL	Netherlands
BE	Belgium	HU	Hungary	NO	Norway
BF	Burkina Faso	IE	Iceland	NZ	New Zealand
BG	Bulgaria	IT	Italy	PL	Poland
BJ	Benin	JP	Japan	PT	Portugal
BR	Brazil	KE	Kenya	RO	Romania
BY	Belarus	KG	Kyrgyzstan	RU	Russian Federation
CA	Canada	KP	Democratic People's Republic of Korea	SD	Sudan
CF	Central African Republic	KR	Republic of Korea	SE	Sweden
CG	Congo	KZ	Kazakhstan	SG	Singapore
CH	Switzerland	LJ	Liechtenstein	SI	Slovenia
CI	Côte d'Ivoire	LK	Sri Lanka	SK	Slovakia
CM	Cameroon	LR	Liberia	SN	Senegal
CN	China	LT	Lithuania	SZ	Swaziland
CS	Czechoslovakia	LU	Luxembourg	TD	Chad
CZ	Czech Republic	LV	Latvia	TG	Togo
DE	Germany	MC	Monaco	TJ	Tajikistan
DK	Denmark	MD	Republic of Moldova	TT	Trinidad and Tobago
EE	Estonia	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	UG	Uganda
FI	Finland	MN	Mongolia	US	United States of America
FR	France	MR	Mauritania	UZ	Uzbekistan
GA	Gabon			VN	Viet Nam

**DETECTION OF A LEPTIN RECEPTOR  
VARIANT AND METHODS FOR REGULATING OBESITY**

5           **1. INTRODUCTION**

The present invention relates to a variant form of the receptor for the *obese* gene product. In particular, the invention relates to methods of detecting this receptor variant in cells and tissues of obese individuals. In 10 addition, it relates to methods of inhibiting or down-regulating expression of this variant in cells to augment their responsiveness to weight regulation by leptin as well as methods of using compounds to directly activate signal transduction pathways associated with this ligand-receptor system.

15

2. **BACKGROUND OF THE INVENTION**

Obesity is not only a nutritional disorder in Western societies, it is also a serious health concern because of its 20 association with adult-onset diabetes, hypertension, and heart disease (Grundy, 1990, *Disease-a-Month* 36:645-696). While there was evidence to suggest that body weight was physiologically regulated, the molecular mechanism has remained elusive. However, animal studies have produced 25 several mouse strains that contain single-gene mutations, resulting in an obese phenotype. One such recessive mutation is manifested in the *ob/ob* mice, and it is referred to as the *obese* (*ob*) mutation.

Zhang et al. (1994, *Nature* 372:425-432) describe the 30 cloning and sequencing of the mouse *ob* gene and its human homolog. When an isolated gene fragment was used as a probe, it was shown to hybridize with RNA only in white adipose tissue by northern blot analysis, but no expression was detected in any other tissue. In addition, the coding 35 sequence of the *ob* gene hybridized to all vertebrate genomic DNAs tested, indicating a high level of conservation of this molecule among vertebrates. The deduced amino acid sequences

are 84% identical between human and mouse, and both molecules contain features of secreted proteins.

In an effort to understand the physiologic function of the *ob* gene, several independent research groups produced recombinant *ob* gene product in bacteria for *in vivo* testing (Pelleymounter et al., 1995, *Science* 269:540-543; Halaas et al., 1995, *Science* 269:543-546; Campfield et al., 1995, *Science* 269:546-549). When the Ob protein (also known as leptin) was injected into grossly obese mice, which possessed two mutant copies of the *ob* gene, the mice exhibited a reduced appetite and began to lose weight. In addition, these studies described a dual action of leptin in both reducing the animals' food intake and in increasing their energy expenditure. Similarly, when normal mice received leptin, they also ate less than the untreated controls. More importantly, Campfield et al. (1995, *Science* 269:546-549) injected leptin directly into lateral ventricle, and observed a reduction in the animals' food intake, suggesting that leptin acts on central neuronal networks to regulate feeding behavior and energy balance. Thus, this result provides evidence that the leptin receptor (also known as OB-R) is expressed by cells in the brain.

Recently, a leptin fusion protein was generated and used to screen for OB-R in a cDNA expression library prepared from mouse choroid plexus, a tissue that lines brain cavities termed ventricles (Tartalia, 1995, *Cell* 83:1263-1271). This approach led to the cloning of one form of the OB-R coding sequence, which reveals a single membrane-spanning receptor, sharing structural similarities with several Class I cytokine receptors, such as the gp130 signal-transducing component of the interleukin-6 receptor (Taga et al., 1989, *Cell* 58:573-581), the granulocyte-colony stimulating factor receptor (Fukunaga et al., 1990, *Cell* 61:341-350), and the leukemia inhibitory factor receptor (Gearing et al., 1991, *EMBO J.* 10:2839-2848). Northern blot analysis and reverse transcription-polymerase chain reaction (RT-PCR) demonstrate

that OB-R mRNA is expressed in several tissues, including lung, kidney, total brain, choroid plexus and hypothalamus.

The reported mouse OB-R protein contains a relatively short intracellular cytoplasmic domain as compared with other 5 Class I cytokine receptors. Subsequently, when cDNA encoding its human homolog was isolated from a human infant brain library, the predicted human protein sequence contains a much longer intracellular domain. In view of this finding, it was speculated that different forms of the receptor might exist 10 (Barinaga, 1996, *Science* 271:29). However, prior to the present invention, there was no report on the identification of any variant forms of the OB-R in humans or how such molecules, if they exist, would relate to obesity.

Additionally, several studies have shown that ob gene 15 expression is actually increased in obese humans (Considine et al., 1995, *J. Clin. Invest.* 95:2986-2988; Lonnquist et al., 1995, *Nature Med.* 1:950; Hamilton et al., 1995, *Nature Med.* 1:953). Moreover, the mutations in the mouse *Ob* gene were not detected in human mRNA. Therefore, taken 20 collectively, these studies imply that decreased leptin levels are not the primary cause of obesity, and argue for the presence of a less responsive receptor in obese individuals. There remains a need to isolate such an OB-R variant for the design of therapeutics to augment weight 25 regulation by leptin.

### 3. SUMMARY OF THE INVENTION

The present invention relates to a variant form of the human OB-R. In particular, it relates to the detection of 30 this receptor variant in cells of obese individuals, and methods for treating obesity by targeting this variant.

The invention is based, in part, upon the Applicants' discovery of human cDNA clones encoding a variant form of the OB-R. This receptor differs structurally from a reported 35 OB-R with only three amino acid substitutions in the extracellular domain, but extensive diversity is observed in their intracellular cytoplasmic domains at the 3' end. The

cytoplasmic domain of the variant of the invention is both shorter and distinct in nucleotide sequence from the corresponding domain of the published form of OB-R. Therefore, a wide variety of uses are encompassed by the 5 present invention, including but not limited to, the detection of the receptor variant in cells of obese individuals, methods to inhibit and/or down-regulate the expression of this receptor variant, gene therapy to replace the receptor variant in homozygous individuals, and direct 10 activation of downstream signal transduction pathways in cells expressing the receptor variant for weight regulation.

#### 4. BRIEF DESCRIPTION OF THE DRAWINGS

Figure 15 1A-1E. Nucleotide sequence (SEQ ID NO:1) and deduced amino acid sequence (SEQ ID NO: 2, 3 and 4) of the human OB-R variant. The amino acid sequence diverges from the human OB-R reported by Tartaglia et al. (1995, Cell 83:1263-1271) at nucleotide residue #349, #422, #764 and from residue #2770 and beyond.

20

#### 5. DETAILED DESCRIPTION OF THE INVENTION

##### 5.1. THE OB-R VARIANT

The present invention relates to nucleic acid and amino acid sequences of an OB-R variant in the Class I cytokine 25 receptor family. In a specific embodiment by way of example in Section 6, infra, this variant was cloned and characterized. Amino acid sequence comparison of this OB-R variant with a published human OB-R sequence (Tartaglia et al., 1995, Cell 83:1263-1271) reveals three amino acid 30 differences in their extracellular domain and extensive diversity in their intracellular cytoplasmic domains. More specifically, Figure 1A-1E shows that in the variant, nucleotide residues #349-351 encode alanine, nucleotide residues #421-423 encode arginine and nucleotide residues 35 #763-765 encode arginine. Additionally, the variant diverges both in length and sequence composition from the published human OB-R sequence from nucleotide residue #2770 and beyond.

In order to clone additional variant forms of the molecule, labeled DNA probes made from nucleic acid fragments corresponding to any portion of the cDNA disclosed herein may be used to screen a cDNA library prepared from human fetal liver, human lung, human kidney, human choroid plexus and human hypothalamus. More specifically, oligonucleotides corresponding to either the 5' or 3' terminus of the cDNA sequence may be used to obtain longer nucleotide sequences.

Briefly, the library may be plated out to yield a maximum of 30,000 pfu for each 150 mm plate. Approximately 40 plates may be screened. The plates are incubated at 37°C until the plaques reach a diameter of 0.25 mm or are just beginning to make contact with one another (3-8 hours). Nylon filters are placed onto the soft top agarose and after 60 seconds, the filters are peeled off and floated on a DNA denaturing solution consisting of 0.4N sodium hydroxide. The filters are then immersed in neutralizing solution consisting of 1M Tris HCl, pH 7.5, before being allowed to air dry. The filters are prehybridized in casein hybridization buffer containing 10% dextran sulfate, 0.5M NaCl, 50mM Tris HCl, pH 7.5, 0.1% sodium pyrophosphate, 1% casein, 1% SDS, and denatured salmon sperm DNA at 0.5 mg/ml for 6 hours at 60°C. The radiolabelled probe is then denatured by heating to 95°C for 2 minutes and then added to the prehybridization solution containing the filters. The filters are hybridized at 60°C for 16 hours. The filters are then washed in 1X wash mix (10X wash mix contains 3M NaCl, 0.6M Tris base, and 0.02M EDTA) twice for 5 minutes each at room temperature, then in 1X wash mix containing 1% SDS at 60°C for 30 minutes, and finally in 0.3X wash mix containing 0.1% SDS at 60°C for 30 minutes. The filters are then air dried and exposed to x-ray film for autoradiography. After developing, the film is aligned with the filters to select a positive plaque. If a single, isolated positive plaque cannot be obtained, the agar plug containing the plaques will be removed and placed in lambda dilution buffer containing 0.1M NaCl, 0.01M magnesium sulfate, 0.035M Tris HCl, pH 7.5, 0.01% gelatin. The phage

may then be replated and rescreened to obtain single, well isolated positive plaques. Positive plaques may be isolated and the cDNA clones sequenced using primers based on the known cDNA sequence. This step may be repeated until a full 5 length cDNA is obtained.

One method for identifying all 3' isoforms is to PCR amplify the 3' ends of the variant cDNA from a variety of tissues including but not limiting to, choroid plexus, hypothalamus, fetal liver, bone marrow, ovary, or prostate. 10 To obtain the 3' end of the cDNA, an oligo-dT primer is used to synthesize the cDNA first strand. OB-R specific primers from the conserved region of the gene (e.g. up stream of nucleotide 2770) and oligo-dT are then used to amplify the 3' end. The PCR fragments are cloned and sequenced by standard 15 techniques. Once obtained, these sequences may be translated into amino acid sequence and examined for certain landmarks such as continuous open reading frame, regulatory regions that associate with tyrosine kinase activation, and finally overall structural similarity to known OB-R variants.

20

#### 5.2. EXPRESSION OF THE OB-R VARIANT

In accordance with the invention, the OB-R variant polynucleotide sequence which encodes a protein, peptide fragments, fusion proteins or functional equivalents thereof, 25 may be used to generate recombinant DNA molecules that direct the expression of the protein, peptide fragments, fusion proteins or a functional equivalent thereof, in appropriate host cells. Such polynucleotide sequences, as well as other polynucleotides which selectively hybridize to at least a 30 part of such polynucleotides or their complements, may also be used in nucleic acid hybridization assays, Southern and Northern blot analyses, etc.

Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a 35 functionally equivalent amino acid sequence, may be used in the practice of the invention for the expression of the OB-R variant. Such DNA sequences include those which are capable

of hybridizing to the OB-R variant sequence under stringent conditions, particularly at its 3' end. The phrase "stringent conditions" as used herein refers to those hybridizing conditions that (1) employ low ionic strength and 5 high temperature for washing, for example, 0.015 M NaCl/0.0015 M sodium citrate/0.1% SDS at 50°C.; (2) employ during hybridization a denaturing agent such as formamide, for example, 50% (vol/vol) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50 mM sodium 10 phosphate buffer at pH 6.5 with 750 mM NaCl, 75 mM sodium citrate at 42°C; or (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M Sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 g/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC and 15 0.1% SDS.

Altered DNA sequences which may be used in accordance with the invention include deletions, additions or substitutions of different nucleotide residues resulting in a sequence that encodes the same or a functionally equivalent 20 gene product. The gene product itself may contain deletions, additions or substitutions of amino acid residues within the OB-R variant sequence, which result in a silent change thus producing a functionally equivalent protein. Such amino acid 25 substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine, histidine and arginine; amino acids with uncharged 30 polar head groups having similar hydrophilicity values include the following: glycine, asparagine, glutamine, serine, threonine, tyrosine; and amino acids with nonpolar head groups include alanine, valine, isoleucine, leucine, phenylalanine, proline, methionine, tryptophan.

35 The DNA sequence of the invention may be engineered in order to alter the OB-R variant coding sequence for a variety of ends, including but not limited to, alterations which

modify processing and expression of the gene product. For example, mutations may be introduced using techniques which are well known in the art, e.g., site-directed mutagenesis, to insert new restriction sites, to alter glycosylation patterns, phosphorylation, etc. In addition, the intracellular domain may also be altered and replaced by a different domain, such as the OB-R intracellular domain by Tartaglia et al.

In another embodiment of the invention, the OB-R variant sequence may be ligated to a heterologous sequence to encode a fusion protein. For example, for screening of peptide libraries for inhibitors or stimulators of receptor activity, it may be useful to encode a chimeric protein expressing a heterologous epitope that is recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the OB-R variant sequence and the heterologous protein sequence, so that the variant may be cleaved away from the heterologous moiety.

In an alternate embodiment of the invention, the coding sequence of the OB-R variant could be synthesized in whole or in part, using chemical methods well known in the art. (See, for example, Caruthers et al., 1980, *Nuc. Acids Res. Symp. Ser.* 7:215-233; Crea and Horn, 1980, *Nuc. Acids Res.* 9(10):2331; Matteucci and Caruthers, 1980, *Tetrahedron Letters* 21:719; and Chow and Kempe, 1981, *Nuc. Acids Res.* 9(12):2807-2817). Alternatively, the protein itself could be produced using chemical methods to synthesize OB-R variant amino acid sequence in whole or in part. For example, peptides can be synthesized by solid phase techniques, 30 cleaved from the resin, and purified by preparative high performance liquid chromatography. (e.g., see Creighton, 1983, *Proteins Structures And Molecular Principles*, W.H. Freeman and Co., N.Y. pp. 50-60). The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation procedure; see Creighton, 1983, *Proteins, Structures and Molecular Principles*, W.H. Freeman and Co., N.Y., pp. 34-49).

In order to express the OB-R variant in host cells, the nucleotide sequence coding for the variant, or a functional equivalent, is inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. The expressed gene products as well as host cells or cell lines transfected or transformed with recombinant OB-R variant expression vectors can be used for a variety of purposes. For example, host cells expressing the OB-R variant may be used to verify the ability of this molecule to bind leptin in a binding assay with radiolabeled, enzyme-conjugated or fluorescent dye-conjugated leptin. At the same time, the ability of the molecule to transduce an activation signal in host cells upon binding to leptin may be tested by assaying proliferation or phosphorylation pattern of kinases in the cells. In addition, genetically-engineered host cells can be used to screen for and select agonist and antagonist compounds, including any inhibitors that would interfere with binding of leptin to the extracellular domain of the OB-R variant. In that connection, such host cells may be used to screen for and select small molecules that can supplement the incomplete signal transduced by the OB-R variant following leptin binding. Other uses, include, but are not limited to generating antibodies (i.e., monoclonal or polyclonal) that competitively inhibit activity of an OB-R variant, neutralize its activity, or even enhances its activity. Antibodies may be used in detecting and quantifying expression of OB-R levels in cells and tissues.

30        5.3. USES OF THE OB-R VARIANT POLYNUCLEOTIDE

The OB-R variant polynucleotide may be used for diagnostic and/or therapeutic purposes. For diagnostic purposes, the OB-R variant polynucleotide may be used to detect gene expression or aberrant gene expression in obese individuals as well as in normal individuals to identify predisposition for obesity. Included in the scope of the invention are oligonucleotide sequences, that include

antisense RNA and DNA molecules, ribozymes and triplex DNA, that function to inhibit translation of OB-R variant.

**5.3.1. DIAGNOSTIC USES OF OB-R VARIANT POLYNUCLEOTIDE**

5 The OB-R variant polynucleotide may have a number of uses for the diagnosis of the possible causes underlying obesity, resulting from expression of the receptor variant. For example, the OB-R variant cytoplasmic domain DNA sequence may be used in hybridization assays of biopsies or autopsies 10 to diagnose OB-R variant expression; e.g., Southern or Northern analysis, including *in situ* hybridization assays as well as PCR. Such techniques are well known in the art, and are in fact the basis of many commercially available diagnostic kits. For PCR detection, primers may be designed 15 from a conserved region of the coding sequence and within the 3' region of OB-R variant. The tissues suitable for such analysis include but are not limited to, hypothalamus, choroid plexus, adipose tissues, lung, prostate, ovary, small intestine, bone marrow and peripheral blood mononuclear 20 cells.

**5.3.2. THERAPEUTIC USES OF THE OB-R VARIANT POLYNUCLEOTIDE**

The OB-R variant polynucleotide may be useful in the treatment of various abnormal obese conditions. By 25 introducing gene sequences into cells, gene therapy can be used to treat conditions in which the cells do not respond to leptin normally due to expression of the OB-R variant. In some instances, the polynucleotide encoding a functional OB-R 30 is intended to replace or act in the place of the functionally deficient OB-R variant gene. Alternatively, abnormal conditions characterized by expression of two copies of the OB-R variant can be treated using the gene therapy techniques described below.

35 Non-responsiveness to normal levels of leptin is an important cause of obesity. This may result from a functionally defective receptor that does not transduce

competent signals upon ligand binding. Recombinant gene therapy vectors, such as viral vectors, may be engineered to express signalling competent forms of OB-R which may be used to augment the non-responsiveness of the naturally occurring 5 OB-R variant. A signalling competent form may be, for example, a protein with the same extracellular domain and transmembrane region, but containing all or part of its normal signal transduction domain, such as that described by Tartaglia et al. (1995, Cell 83:1263-1271). Thus recombinant 10 gene therapy vectors may be used therapeutically for treatment of obesity resulting from expression or activity of the OB-R variant. Accordingly, the invention provides a method of augmenting signal transduction by an endogenous OB-R variant in a cell comprising delivering a DNA molecule 15 encoding a signalling competent form of the OB-R to the cell so that the signalling competent protein is produced in the cell and competes with the endogenous defective OB-R variant for access to molecules in the signalling pathway which does not activate or are not activated by the endogenous natural 20 defective receptor. Additionally, since dimerization of a functional receptor with a defective variant may occur in cells of heterozygous individuals, small molecules may be used to inhibit such pairing, thereby increasing the number of functional dimeric receptors for proper signalling in 25 response to leptin.

In contrast, overexpression of either leptin or a competent OB-R may result in a clinical anorexic-like syndrome due to a loss of appetite or hypermetabolic activity. In such cases, the OB-R variant of the invention 30 may be introduced into cells with functional receptors to cause a decrease in the number of functional receptors or to compete with such receptors for leptin binding.

Expression vectors derived from viruses such as retroviruses, vaccinia virus, adeno-associated virus, herpes 35 viruses, or bovine papilloma virus, may be used for delivery of recombinant functional OB-R into the targeted cell population. Methods which are well known to those skilled in

the art can be used to construct recombinant viral vectors containing an OB-R polynucleotide sequence. See, for example, the techniques described in Sambrook et al., 1989, Molecular Cloning A Laboratory Manual, Cold Spring Harbor 5 Laboratory, N.Y. and Ausubel et al., 1989, Current Protocols in Molecular Biology, Greene Publishing Associates and Wiley Interscience, N.Y. Alternatively, recombinant OB-R molecules can be reconstituted into liposomes for delivery to target cells.

10 Oligonucleotide sequences including anti-sense RNA and DNA molecules and ribozymes that function to inhibit the translation of the OB-R variant mRNA are within the scope of the invention. Anti-sense RNA and DNA molecules act to directly block the translation of mRNA by binding to targeted 15 mRNA and preventing protein translation. In regard to antisense DNA, oligodeoxyribonucleotides derived from the OB-R variant nucleotide sequence at nucleotide #2771 and beyond, are preferred.

Ribozymes are enzymatic RNA molecules capable of 20 catalyzing the specific cleavage of RNA. The mechanism of ribozyme action involves sequence specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. Within the scope of the invention are engineered hammerhead motif ribozyme molecules 25 that specifically and efficiently catalyze endonucleolytic cleavage of OB-R variant RNA sequences.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites which include the 30 following sequences, GUA, GUU and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the target gene containing the cleavage site may be evaluated for predicted structural features such as secondary structure that may render the 35 oligonucleotide sequence unsuitable. The suitability of candidate targets may also be evaluated by testing their

accessibility to hybridization with complementary oligonucleotides, using ribonuclease protection assays.

Oligodeoxyribonucleotides can form sequence-specific triple helices by hydrogen bonding to specific complementary sequences in duplexed DNA. Interest in triple helices has focused on the potential biological and therapeutic applications of these structures. Formation of specific triple helices may selectively inhibit the replication and/or gene expression of targeted genes by prohibiting the specific binding of functional trans-acting factors.

Oligonucleotides to be used in triplex helix formation should be single stranded and composed of deoxynucleotides. The base composition of these oligonucleotides must be designed to promote triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of either purines or pyrimidines to be present on one strand of a duplex. Oligonucleotide sequences may be pyrimidine-based, which will result in TAT and CGC triplets across the three associated strands of the resulting triple helix. The pyrimidine-rich oligonucleotides provide base complementarity to a purine-rich region of a single strand of the duplex in a parallel orientation to that strand. In addition, oligonucleotides may be chosen that are purine-rich, for example, containing a stretch of G residues. These oligonucleotides will form a triple helix with a DNA duplex that is rich in GC pairs, in which the majority of the purine residues are located on a single strand of the targeted duplex, resulting in GGC triplets across the three strands in the triplex. Alternatively, the potential sequences that can be targeted for triple helix formation may be increased by creating a so called "switchback" oligonucleotide. Switchback oligonucleotides are synthesized in an alternating 5'-3', 3'-5' manner, such that they base pair with first one strand of a duplex and then the other, eliminating the necessity for a sizeable stretch of either purines or pyrimidines to be present on one strand of a duplex.

Both anti-sense RNA and DNA molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of RNA molecules. These include techniques for chemically synthesizing oligodeoxyribonucleotides well known in the art such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA sequences encoding the antisense RNA molecule. Such DNA sequences may be incorporated into a wide variety of vectors which incorporate suitable RNA polymerase promoters such as the T7 or SP6 polymerase promoters. Alternatively, antisense cDNA constructs that synthesize antisense RNA constitutively or inducibly, depending on the promoter used, can be introduced stably into cell lines.

15 Various modifications to the DNA molecules may be introduced as a means of increasing intracellular stability and half-life. Possible modifications include but are not limited to the addition of flanking sequences of ribo- or deoxy- nucleotides to the 5' and/or 3' ends of the molecule 20 or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the oligodeoxyribonucleotide backbone.

#### 5.4. ACTIVATION OF TYROSINE KINASE PATHWAYS IN OBESITY

25 Many known class I cytokine receptors initiate cell signaling via Janus kinases (JAKs) (Ihle, 1995, *Nature* 377:591-594; Heldin, 1995, *Cell* 80:213-223; Kishimoto et al., 1994, *Cell* 76:253-62; Ziemiczki et al., 1994, *Trends Cell. Biol.* 4:207-212). JAK1-3 have been shown to bind to conserved 30 sequences termed box1 and box2 (Fukunaga et al., 1991, *EMBO J.* 10:2855-65; Murakami, 1991, *Proc. Natl. Acad. Sci. USA* 88:11349-53). Ligand binding induces a homo- or hetero-dimerization of receptor chains which activates, by phosphorylation, the JAKs. The activated JAKs, in turn, 35 phosphorylate members of the STAT family (Heldin, 1995, *Cell* 80:213-223; Kishimoto et al., *Blood* 86:1243-54; Darnell et al., 1994, *Science* 264:1415-21; Zhong et al., 1994, *Proc.*

Natl. Acad. Sci. USA 91:4806-10; Hou et al., 1994, Science 265:1701-6). These phosphorylated STATs ultimately translocate to the nucleus, form transcription complexes, and regulate gene expression. Both box1 and box2 are required 5 for complete signaling in certain systems. (Fukunaga et al., 1991, EMBO J. 10:2855-65; Murakami, 1991, Proc. Natl. Acad. Sci. USA 88:11349-53). The OB-R variant disclosed herein has a typical box1 (from nucleotide #2707-2730) that contains the critical xWxxxPxP amino acid sequence, but it does not 10 contain an obvious box2 nor further downstream sequences that are important for normal receptor activation. Therefore, it is possible to use compounds that activate JAKs to directly activate these pathways for weight regulation without triggering the OB-R.

15

#### 6. EXAMPLE: MOLECULAR CLONING OF AN OB-R VARIANT

A number of cDNA clones were isolated from a human fetal liver cDNA library (Clontech, Palo Alto, CA), and the DNA sequences of several of these clones were determined. These 20 clones (designated as Hu-B1.219 #4, #33, #34, #1, #8) contained overlapping sequences, which were then compiled into a contiguous nucleotide sequence (Figure 1A-1E). When the deduced amino acid sequence of one such sequence was compared with the sequence of a recently published human 25 OB-R, they were shown to be nearly identical in the extracellular domains with the exception of three amino acids, whereas there existed extensive diversity in their intracellular cytoplasmic domains at the 3' end. The predicted protein sequence contains two FN III domains, each 30 containing a "WS box", which are characteristic of genes of the Class I cytokine receptor family. Therefore, the cDNA disclosed herein encodes an OB-R variant.

When various human tissue RNA were probed with a fragment of this OB-R variant by Northern blot analysis, 35 expression of this molecule was detected in heart, placenta, lung, liver, muscle, pancreas, prostate, ovary, small intestine and brain.

Based on the sequence presented in Figure 1A-1E, the translation initiation site appears at position #97. The sequence encodes an open reading frame up to and including nucleotide #2814. It is believed that the sequence between 5 nucleotides #2629 and #2682 encodes a transmembrane domain. The complete sequence encodes a protein of 906 amino acids.

The sequence of the OB-R variant is identical to the sequence of human OB-R reported by Tartaglia (1995, Cell 83:1263-1271) in the transmembrane region and a portion of 10 the intracellular domain up to and including nucleotide #2769, then they diverge at nucleotide #2770 and beyond. In addition, the product of this cDNA is substantially shorter in its intracellular domain than the published human OB-R. These two forms of OB-R may derive from a common precursor 15 mRNA by an alternative splicing mechanism. The sequence in this region is consistent with well known splice junctions.

#### 7. DEPOSIT OF MICROORGANISMS

The following organisms were deposited with the American 20 Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852.

Strain Designation Accession No.

HuB1.219, #1	75885
HuB1.219, #4	75886
HuB1.219, #33	75888
HuB1.219, #34	75889
HuB1.219, #8	75974

The present invention is not to be limited in scope by the exemplified embodiments, which are intended as 30 illustrations of individual aspects of the invention.

Indeed, various modifications for the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and accompanying drawings. Such modifications are intended to 35 fall within the scope of the appended claims.

All publications cited herein are incorporated by reference in their entirety.

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

(i) APPLICANT: Snodgrass, H.  
 Cioffi, Joseph  
 Zupancic, Thomas  
 Shafer, Alan

(ii) TITLE OF INVENTION: DETECTION OF A LEPTIN RECEPTOR VARIANT  
 AND METHODS FOR REGULATING OBESITY

(iii) NUMBER OF SEQUENCES: 4

## (iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Pennie & Edmonds  
 (B) STREET: 1155 Avenue of the Americas  
 (C) CITY: New York  
 (D) STATE: New York  
 (E) COUNTRY: US  
 (F) ZIP: 10036-2711

## (v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk  
 (B) COMPUTER: IBM PC compatible  
 (C) OPERATING SYSTEM: PC-DOS/MS-DOS  
 (D) SOFTWARE: PatentIn Release #1.0, Version #1.30

## (vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: US 08/588,526  
 (B) FILING DATE: 18-JAN-1996  
 (C) CLASSIFICATION:

## (vii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Poissant, Brian M.  
 (B) REGISTRATION NUMBER: 28,462  
 (C) REFERENCE/DOCKET NUMBER: 8907-030

## (ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (212) 790-9090  
 (B) TELEFAX: (212) 869-9741  
 (C) TELEX: 66141 PENNIE

## (2) INFORMATION FOR SEQ ID NO:1:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2880 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA

## (ix) FEATURE:

(A) NAME/KEY: CDS  
 (B) LOCATION: 1..2880

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GCG CGC GCG ACG CAG GTG CCC GAG CCC CGG CCC GCG CCC ATC TCT GCC  
 Ala Arg Ala Thr Gln Val Pro Glu Pro Arg Pro Ala Pro Ile Ser Ala  
 1 5 10 15

48

TTC GGT CGA GTT GGA CCC CCG GAT CAA GGT GTA CTT CTC TGA AGT AAG

96

Phe Gly Arg Val Gly Pro Pro Asp Gln Gly Val Leu Leu \* Ser Lys  
 20 25 30

ATG ATT TGT CAA AAA TTC TGT GTG GTT TTG TTA CAT TGG GAA TTT ATT 144  
 Met Ile Cys Gln Lys Phe Cys Val Val Leu Leu His Trp Glu Phe Ile  
 35 40 45

TAT GTG ATA ACT GCG TTT AAC TTG TCA TAT CCA ATT ACT CCT TGG AGA 192  
 Tyr Val Ile Thr Ala Phe Asn Leu Ser Tyr Pro Ile Thr Pro Trp Arg  
 50 55 60

TTT AAG TTG TCT TGC ATG CCA CCA AAT TCA ACC TAT GAC TAC TTC CTT 240  
 Phe Lys Leu Ser Cys Met Pro Pro Asn Ser Thr Tyr Asp Tyr Phe Leu  
 65 70 75 80

TTG CCT GCT GGA CTC TCA AAG AAT ACT TCA AAT TCG AAT GGA CAT TAT 288  
 Leu Pro Ala Gly Leu Ser Lys Asn Thr Ser Asn Ser Asn Gly His Tyr  
 85 90 95

GAG ACA GCT GTT GAA CCT AAG TTT AAT TCA AGT GGT ACT CAC TTT TCT 336  
 Glu Thr Ala Val Glu Pro Lys Phe Asn Ser Ser Gly Thr His Phe Ser  
 100 105 110

AAC TTA TCC AAA GCA ACT TTC CAC TGT TGC TTT CGG AGT GAG CAA GAT 384  
 Asn Leu Ser Lys Ala Thr Phe His Cys Cys Phe Arg Ser Glu Gln Asp  
 115 120 125

AGA AAC TGC TCC TTA TGT GCA GAC AAC ATT GAA GGA AGG ACA TTT GTT 432  
 Arg Asn Cys Ser Leu Cys Ala Asp Asn Ile Glu Gly Arg Thr Phe Val  
 130 135 140

TCA ACA GTA AAT TCT TTA GTT TTT CAA CAA ATA GAT GCA AAC TGG AAC 480  
 Ser Thr Val Asn Ser Leu Val Phe Gln Gln Ile Asp Ala Asn Trp Asn  
 145 150 155 160

ATA CAG TGC TGG CTA AAA GGA GAC TTA AAA TTA TGC ATC TGT TAT GTG 528  
 Ile Gln Cys Trp Leu Lys Gly Asp Leu Lys Leu Phe Ile Cys Tyr Val  
 165 170 175

GAG TCA TTA TTT AAG AAT CTA TTC AGG AAT TAT AAC TAT AAG GTC CAT 576  
 Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn Tyr Lys Val His  
 180 185 190

CTT TTA TAT GTT CTG CCT GAA GTG TTA GAA GAT TCA CCT CTG GTT CCC 624  
 Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser Pro Leu Val Pro  
 195 200 205

CAA AAA GGC AGT TTT CAG ATG GTT CAC TGC AAT TGC AGT GTT CAT GAA 672  
 Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys Ser Val His Glu  
 210 215 220

TGT TGT GAA TGT CTT GTG CCT GTG CCA ACA GCC AAA CTC AAC GAC ACT 720  
 Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys Leu Asn Asp Thr  
 225 230 235 240

CTC CTT ATG TGT TTG AAA ATC ACA TCT GGT GGA GTA ATT TTC CGG TCA 768  
 Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val Ile Phe Arg Ser  
 245 250 255

CCT CTA ATG TCA GTT CAG CCA ATA AAT ATG GTG AAG CCT GAT CCA CCA 816  
 Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys Pro Asp Pro Pro  
 260 265 270

TTA GGT TTG CAT ATG GAA ATC ACA GAT GAT GGT AAT TTA AAG ATT TCT 864  
 Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn Leu Lys Ile Ser  
 275 280 285

TGG TCC AGC CCA-CCA TTG GTA CCA TTT CCA CTT CAA TAT CAA GTG AAA Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln Tyr Gln Val Lys 290 295 300	912
TAT TCA GAG AAT TCT ACA ACA GTT ATC AGA GAA GCT GAC AAG ATT GTC Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala Asp Lys Ile Val 305 310 315 320	960
TCA GCT ACA TCC CTG CTA GTA GAC AGT ATA CTT CCT GGG TCT TCG TAT Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro Gly Ser Ser Tyr 325 330 335	1008
GAG GTT CAG GTG AGG GGC AAG AGA CTG GAT GGC CCA GGA ATC TGG AGT Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro Gly Ile Trp Ser 340 345 350	1056
GAC TGG AGT ACT CCT CGT GTC TTT ACC ACA CAA GAT GTC ATA TAC TTT Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp Val Ile Tyr Phe 355 360 365	1104
CCA CCT AAA ATT CTG ACA AGT GTT GGG TCT AAT GTT TCT TTT CRC TGC Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val Ser Phe His Cys 370 375 380	1152
ATC TAT AAG AAG GAA AAC AAG ATT GTT CCC TCA AAA GAG ATT GTT TGG Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys Glu Ile Val Trp 385 390 395 400	1200
TGG ATG AAT TTA GCT GAG AAA ATT CCT CAA AGC CAG TAT GAT GTT GTG Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln Tyr Asp Val Val 405 410 415	1248
AGT GAT CAT GTT AGC AAA GTT ACT TTT TTC AAT CTG AAT GAA ACC AAA Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu Asn Glu Thr Lys 420 425 430	1296
CCT CGA GGA AAG TTT ACC TAT GAT GCA GTG TAC TGC TGC AAT GAA CAT Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys Cys Asn Glu His 435 440 445	1344
GAA TGC CAT CAT CGC TAT GCT GAA TTA TAT GTG ATT GAT GTC AAT ATC Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile Asp Val Asn Ile 450 455 460	1392
AAT ATC TCA TGT GAA ACT GAT GGG TAC TTA ACT AAA ATG ACT TGC AGA Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys Met Thr Cys Arg 465 470 475 480	1440
TGG TCA ACC AGT ACA ATC CAG TCA CTT GCG GAA AGC ACT TTG CAA TTG Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser Thr Leu Gln Leu 485 490 495	1488
AGG TAT CAT AGG AGC AGC CTT TAC TGT TCT GAT ATT CCA TCT ATT CAT Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile Pro Ser Ile His 500 505 510	1536
CCC ATA TCT GAG CCC AAA GAT TGC TAT TTG CAG AGT GAT GGT TTT TAT Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser Asp Gly Phe Tyr 515 520 525	1584
GAA TGC ATT TTC CAG CCA ATC TTC CTA TTA TCT GGC TAC ACA ATG TGG Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly Tyr Thr Met Trp 530 535 540	1632
ATT AGG ATC AAT CAC TCT CTA GGT TCA CTT GAC TCT CCA CCA ACA TGT Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser Pro Pro Thr Cys 545 550 555 560	1680

GTC CTT CCT GAT-TCT GTG GTG AAG CCA CTG CCT CCA TCC AGT GTG AAA Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro Ser Ser Val Lys 565 570 575	1728
CGA GAA ATT ACT ATA AAC ATT GGA TTA TTG AAA ATA TCT TGG GAA AAG Arg Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile Ser Trp Glu Lys 580 585 590	1776
CCA GTC TTT CCA GAG AAT AAC CTT CAA TTC CAG ATT CGC TAT GGT TTA Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile Arg Tyr Gly Leu 595 600 605	1824
AGT GGA AAA GAA GTA CAA TGG AAG ATG TAT GAG GTT TAT GAT CGA AAA Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val Tyr Asp Arg Lys 610 615 620	1872
TCA AAA TCT GTC AGT CTC CCA GTT CCA GAC TTG TGT GCA GTC TAT GCT Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys Ala Val Tyr Ala 625 630 635 640	1920
GTT CAG GTG CGC TGT AAG AGG CTA GAT GGA CTG GGA TAT TGG AGT AAT Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly Tyr Trp Ser Asn 645 650 655	1968
TGG AGC AAT CCA GCC TAC ACA GTT GTC ATG GAT ATA AAA GTT CCT ATG Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile Lys Val Pro Met 660 665 670	2016
AGA GGA CCT GAA TTT TGG AGA ATA ATT AAT GGA GAT ACT ATG AAA AAG Arg Gly Pro Glu Phe Trp Arg Ile Ile Asn Gly Asp Thr Met Lys Lys 675 680 685	2064
GAG AAA AAT GTC ACT TTA CTT TGG AAG CCC CTG ATG AAA AAT GAC TCA Glu Lys Asn Val Thr Leu Leu Trp Lys Pro Leu Met Lys Asn Asp Ser 690 695 700	2112
TTG TGC AGT GTT CAG AGA TAT GTG ATA AAC CAT CAT ACT TCC TGC AAT Leu Cys Ser Val Gln Arg Tyr Val Ile Asn His His Thr Ser Cys Asn 705 710 715 720	2160
GGA ACA TGG TCA GAA GAT GTG GGA AAT CAC ACG AAA TTC ACT TTC CTG Gly Thr Trp Ser Glu Asp Val Gly Asn His Thr Lys Phe Thr Phe Leu 725 730 735	2208
TGG ACA GAG CAA GCA CAT ACT GTT ACG GTT CTG GCC ATC AAT TCA ATT Trp Thr Glu Gln Ala His Thr Val Thr Val Leu Ala Ile Asn Ser Ile 740 745 750	2256
GGT GCT TCT GTT GCA AAT TTT AAT TTA ACC TTT TCA TGG CCT ATG AGC Gly Ala Ser Val Ala Asn Phe Asn Leu Thr Phe Ser Trp Pro Met Ser 755 760 765	2304
AAA GTA AAT ATC GTG CAG TCA CTC AGT GCT TAT CCT TTA AAC AGC AGT Lys Val Asn Ile Val Gln Ser Leu Ser Ala Tyr Pro Leu Asn Ser Ser 770 775 780	2352
TGT GTG ATT GTT TCC TGG ATA CTA TCA CCC AGT GAT TAC AAG CTA ATG Cys Val Ile Val Ser Trp Ile Leu Ser Pro Ser Asp Tyr Lys Leu Met 785 790 795 800	2400
TAT TTT ATT ATT GAG TGG AAA AAT CTT AAT GAA GAT GGT GAA ATA AAA Tyr Phe Ile Ile Glu Trp Lys Asn Leu Asn Glu Asp Gly Glu Ile Lys 805 810 815	2448
TGG CTT AGA ATC TCT TCA TCT GTT AAG AAG TAT TAT ATC CAT GAT CAT Trp Leu Arg Ile Ser Ser Ser Val Lys Lys Tyr Tyr Ile His Asp His 820 825 830	2496

TTT ATC CCC ATT GAG AAG TAC CAG TTC AGT CTT TAC CCA ATA TTT ATG Phe Ile Pro Ile Glu Lys Tyr Gln Phe Ser Leu Tyr Pro Ile Phe Met 835 840 845	2544
GAA GGA GTG GGA AAA CCA AAG ATA ATT AAT AGT TTC ACT CAA GAT GAT Glu Gly Val Gly Lys Pro Lys Ile Ile Asn Ser Phe Thr Gln Asp Asp 850 855 860	2592
ATT GAA AAA CAC CAG AGT GAT GCA GGT TTA TAT GTA ATT GTG CCA GTA Ile Glu Lys His Gln Ser Asp Ala Gly Leu Tyr Val Ile Val Pro Val 865 870 875 880	2640
ATT ATT TCC TCT TCC ATC TTA TTG CTT GGA ACA TTA TTA ATA TCA CAC Ile Ile Ser Ser Ile Leu Leu Leu Gly Thr Leu Leu Ile Ser His 885 890 895	2688
CAA AGA ATG AAA AAG CTA TTT TGG GAA GAT GTT CCG AAC CCC AAG AAT Gln Arg Met Lys Lys Leu Phe Trp Glu Asp Val Pro Asn Pro Lys Asn 900 905 910	2736
TGT TCC TGG GCA CAA GGA CCT AAT TTT CAG AAG AAA ATG CCT GGC ACA Cys Ser Trp Ala Gln Gly Pro Asn Phe Gln Lys Lys Met Pro Gly Thr 915 920 925	2784
AAG GAA CTA CTG GGT GGA GGT TGG TTG ACT TAG GAA ATG CTT GTG AAG Lys Glu Leu Leu Gly Gly Trp Leu Thr * Glu Met Leu Val Lys 930 935 940	2832
CTA CGT CCT ACC TCG TGC GCA CCT GCT CTC CCT GAG GTG TGC ACA ATG Leu Arg Pro Thr Ser Cys Ala Pro Ala Leu Pro Glu Val Cys Thr Met 945 950 955 960	2880

## (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 29 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Ala Arg Ala Thr Gln Val Pro Glu Pro Arg Pro Ala Pro Ile Ser Ala 1 5 10 15
Phe Gly Arg Val Gly Pro Pro Asp Gln Gly Val Leu Leu 20 25

## (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 908 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Ser Lys Met Ile Cys Gln Lys Phe Cys Val Val Leu Leu His Trp Glu 1 5 10 15
Phe Ile Tyr Val Ile Thr Ala Phe Asn Leu Ser Tyr Pro Ile Thr Pro

20 -	25	30
Trp Arg Phe Lys Leu Ser Cys Met Pro Pro Asn Ser Thr Tyr Asp Tyr 35	40	45
Phe Leu Leu Pro Ala Gly Leu Ser Lys Asn Thr Ser Asn Ser Asn Gly 50	55	60
His Tyr Glu Thr Ala Val Glu Pro Lys Phe Asn Ser Ser Gly Thr His 65	70	75
Phe Ser Asn Leu Ser Lys Ala Thr Phe His Cys Cys Phe Arg Ser Glu 85	90	95
Gln Asp Arg Asn Cys Ser Leu Cys Ala Asp Asn Ile Glu Gly Arg Thr 100	105	110
 Phe Val Ser Thr Val Asn Ser Leu Val Phe Gln Gln Ile Asp Ala Asn 115		
120		
125		
Trp Asn Ile Gln Cys Trp Leu Lys Gly Asp Leu Lys Leu Phe Ile Cys 130	135	140
Tyr Val Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn Tyr Lys 145	150	155
Val His Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser Pro Leu 165	170	175
Val Pro Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys Ser Val 180	185	190
His Glu Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys Leu Asn 195	200	205
Asp Thr Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val Ile Phe 210	215	220
Arg Ser Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys Pro Asp 225	230	235
Pro Pro Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn Leu Lys 245	250	255
Ile Ser Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln Tyr Gln 260	265	270
Val Lys Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala Asp Lys 275	280	285
Ile Val Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro Gly Ser 290	295	300
Ser Tyr Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro Gly Ile 305	310	315
Trp Ser Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp Val Ile 325	330	335
Tyr Phe Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val Ser Phe 340	345	350
His Cys Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys Glu Ile 355	360	365

Val Trp Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln Tyr Asp  
 370 375 380  
 Val Val Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu Asn Glu  
 385 390 395 400  
 Thr Lys Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys Cys Asn  
 405 410 415  
 Glu His Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile Asp Val  
 420 425 430  
 Asn Ile Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys Met Thr  
 435 440 445  
 Cys Arg Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser Thr Leu  
 450 455 460  
 Gln Leu Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile Pro Ser  
 465 470 475 480  
 Ile His Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser Asp Gly  
 485 490 495  
 Phe Tyr Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly Tyr Thr  
 500 505 510  
 Met Trp Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser Pro Pro  
 515 520 525  
 Thr Cys Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro Ser Ser  
 530 535 540  
 Val Lys Arg Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile Ser Trp  
 545 550 555 560  
 Glu Lys Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile Arg Tyr  
 565 570 575  
 Gly Leu Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val Tyr Asp  
 580 585 590  
 Arg Lys Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys Ala Val  
 595 600 605  
 Tyr Ala Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly Tyr Trp  
 610 615 620  
 Ser Asn Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile Lys Val  
 625 630 635 640  
 Pro Met Arg Gly Pro Glu Phe Trp Arg Ile Ile Asn Gly Asp Thr Met  
 645 650 655  
 Lys Lys Glu Lys Asn Val Thr Leu Leu Trp Lys Pro Leu Met Lys Asn  
 660 665 670  
 Asp Ser Leu Cys Ser Val Gln Arg Tyr Val Ile Asn His His Thr Ser  
 675 680 685  
 Cys Asn Gly Thr Trp Ser Glu Asp Val Gly Asn His Thr Lys Phe Thr  
 690 695 700  
 Phe Leu Trp Thr Glu Gln Ala His Thr Val Thr Val Leu Ala Ile Asn  
 705 710 715 720  
 Ser Ile Gly Ala Ser Val Ala Asn Phe Asn Leu Thr Phe Ser Trp Pro

725	730	735
Met Ser Lys Val Asn Ile Val Gln Ser Leu Ser Ala Tyr Pro Leu Asn 740	745	750
Ser Ser Cys Val Ile Val Ser Trp Ile Leu Ser Pro Ser Asp Tyr Lys 755	760	765
Leu Met Tyr Phe Ile Ile Glu Trp Lys Asn Leu Asn Glu Asp Gly Glu 770	775	780
Ile Lys Trp Leu Arg Ile Ser Ser Val Lys Lys Tyr Tyr Ile His 785	790	795
Asp His Phe Ile Pro Ile Glu Lys Tyr Gln Phe Ser Leu Tyr Pro Ile 805	810	815
Phe Met Glu Gly Val Gly Lys Pro Lys Ile Ile Asn Ser Phe Thr Gln 820	825	830
Asp Asp Ile Glu Lys His Gln Ser Asp Ala Gly Leu Tyr Val Ile Val 835	840	845
Pro Val Ile Ile Ser Ser Ile Leu Leu Leu Gly Thr Leu Leu Ile 850	855	860
Ser His Gln Arg Met Lys Lys Leu Phe Trp Glu Asp Val Pro Asn Pro 865	870	875
Lys Asn Cys Ser Trp Ala Gln Gly Pro Asn Phe Gln Lys Lys Met Pro 885	890	895
Gly Thr Lys Glu Leu Leu Gly Gly Trp Leu Thr 900	905	

## (2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 21 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Glu	Met	Leu	Val	Lys	Leu	Arg	Pro	Thr	Ser	Cys	Ala	Pro	Ala	Leu	Pro
1				5				10				15			
Glu Val Cys Thr Met															
20															

International Application No: PCT/ /

<b>MICROORGANISMS</b>	
Optional Sheet in connection with the microorganism referred to on page <u>16</u> , lines <u>17-37</u> , of the description	
<b>A. IDENTIFICATION OF DEPOSIT*</b>	
Further deposits are identified on an additional sheet	
Name of depositary institution American Type Culture Collection	
Address of depositary institution (including postal code and country) 12301 Parklawn Drive Rockville, MD 20852 US	
Date of deposit * September 14, 1994 Accession Number * 75885	
<b>B. ADDITIONAL INDICATIONS*</b> (Leave blank if not applicable). This information is contained on a separate attached sheet	
<b>C. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE*</b> (or information can be obtained therefrom)	
<b>D. SEPARATE FURNISHING OF INDICATIONS*</b> (Leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later *(Specify the general nature of the indications e.g., "Accession Number of Deposit")	
<b>E. <input type="checkbox"/> This sheet was received with the International application when filed (to be checked by the receiving Office)</b>	
(Authorized Officer)	
<input type="checkbox"/> The date of receipt (from the applicant) by the International Bureau *	
WAS	
(Authorized Officer)	

Form PCT/RD/134 (January 1981)

International Application No: PCT/

Form PCT/RO/134 (cont.)

American Type Culture Collection

12301 Parklawn Drive  
Rockville, MD 20852  
US

<u>Accession No.</u>	<u>Date of Deposit</u>
75886	September 14, 1994
75888	September 14, 1994
75889	September 14, 1994
75974	December 14, 1994

WHAT IS CLAIMED IS:

1. A method for detecting a defective OB-R in cells comprising:

- 5           (a) extracting RNA from a cell population;  
             (b) contacting the RNA with an oligonucleotide derived from a portion of the sequence depicted in Figure 1A-1E; and  
             (c) detecting hybridization of the RNA with the oligonucleotide.

10

2. The method of Claim 1 in which the cell population is obtained from the brain.

15           3. The method of Claim 1 in which the cell population is obtained from the lung.

4. The method of Claim 1 in which the cell population is obtained from the kidney.

20           5. The method of Claim 1 in which the oligonucleotide is derived from nucleotide residue #2770 and beyond in the sequence depicted in Figure 1A-1E.

25           6. A method for treating obesity, comprising administering to an individual an effective amount of an agent capable of inhibiting expression of an OB-R variant gene.

30           7. The method of Claim 6 in which the OB-R variant gene further comprises the sequence of Figure 1A-1E or which is capable of selectively hybridizing to it.

35           8. The method of Claim 7 in which the agent is an antisense molecule complementary to mRNA encoded by the sequence of Figure 1A-1E.

9. The method of Claim 7 in which the agent is a ribozyme molecule specific for mRNA encoded by the sequence of Figure 1A-1E.

5 10. The method of Claim 7 in which the agent is a triple helix component.

11. A method for identifying a compound capable of supplementing biological activity of leptin, comprising:  
10 (a) incubating host cell expressing an OB-R variant with leptin;  
(b) incubating a portion of the leptin-treated cells with a test compound; and  
(c) comparing activation signal in the cells  
15 treated in step (b) with cells treated in step (a);

thereby determining whether the compound augments activation of the OB-R variant by leptin.

20 12. The method of Claim 11 in which the OB-R variant is encoded by the sequence depicted in Figure 1A-1E.

25

30

35

9	18	27	36	45	54												
GCG	CGC	GCG	ACG	CAG	GTG	CCC	GAG	CCC	CGG	CCC	GCG	CCC	ATC	TCT	GCC	TTC	GGT
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
A	R	A	T	Q	V	P	E	P	R	P	A	P	I	S	A	F	G
63	72	81	90	99	108												
CGA	GTT	GGA	CCC	CCG	GAT	CAA	GGT	GTA	CTT	CTC	TGA	AGT	AAG	ATG	ATT	TGT	CAA
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
R	V	G	P	P	D	Q	G	V	L	L	*	S	K	M	I	C	Q
117	126	135	144	153	162												
AAA	TTC	TGT	GTG	GTT	TTG	TTA	CAT	TOG	GAA	TTT	ATT	TAT	GTG	ATA	ACT	GCG	TTT
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
K	F	C	V	V	L	L	H	W	E	F	I	Y	V	I	T	A	F
171	180	189	198	207	216												
AAC	TTG	TCA	TAT	CCA	ATT	ACT	CCT	TGG	AGA	TTT	AAG	TTG	TCT	TGC	ATG	CCA	CCA
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
N	L	S	Y	P	I	T	P	W	R	F	K	L	S	C	M	P	P
225	234	243	252	261	270												
AAT	TCA	ACC	TAT	GAC	TAC	TTC	CTT	TTG	CCT	GCT	GGA	CTC	TCA	AAG	ATT	ACT	TCA
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
N	S	T	Y	D	Y	F	L	L	P	A	G	L	S	K	N	T	S
279	288	297	306	315	324												
AAT	TCG	AAT	GGA	CAT	TAT	GAG	ACA	GCT	GTT	GAA	CCT	AAG	TTT	AAT	TCA	AGT	GGT
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
N	S	N	G	H	Y	E	T	A	V	E	P	K	F	N	S	S	G
333	342	351	360	369	378												
ACT	CAC	TTT	TCT	AAC	TTA	TCC	AAA	GCA	ACT	TTC	CAC	TGT	TGC	TTT	CGG	AGT	GAG
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
T	H	F	S	N	L	S	K	A	T	F	H	C	C	F	R	S	E
387	396	405	414	423	432												
CAA	GAT	AGA	AAC	TGC	TCC	TTA	TGT	GCA	GAC	AAC	ATT	GAA	GGA	AGG	ACA	TTT	GTT
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Q	D	R	N	C	S	L	C	A	D	N	I	E	G	R	T	F	V
441	450	459	468	477	486												
TCA	ACA	GTA	AAT	TCT	TTA	GTT	TTT	CAA	CAA	ATA	GAT	GCA	AAC	TGG	AAC	ATA	CAG
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
S	T	V	N	S	L	V	F	Q	Q	I	D	A	N	W	N	I	Q
495	504	513	522	531	540												
TGC	TGG	CTA	AAA	GGA	GAC	TTA	AAA	TTA	TTC	ATC	TGT	TAT	GTG	GAG	TCA	TTA	TTT
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
C	W	L	K	G	D	L	K	L	F	I	C	Y	V	E	S	L	F
549	558	567	576	585	594												
AAG	AAT	CTA	TTC	AGG	AAT	TAT	AAC	TAT	AAG	GTC	CAT	CTT	TTA	TAT	GTT	CTG	CCT
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
K	N	L	F	R	N	Y	N	Y	K	V	H	L	L	Y	V	L	P
603	612	621	630	639	648												
GAA	GTG	TTA	GAA	GAT	TCA	CCT	CTG	GTG	CCC	CAA	AAA	GGC	AGT	TTT	CAG	ATG	GTT
-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
E	V	L	E	D	S	P	L	V	P	Q	K	G	S	F	Q	M	V

Figure 1 A

657	666	675	684	693	702
CAC TGC AAT TGC AGT GTT CAT GAA TGT TGT GAA TGT CTT GTG CCT GTG CCA ACA					
H C N C S V H E C C E C L V P V P T					
711	720	729	738	747	756
GCC AAA CTC AAC GAC ACT CTC CTT ATG TGT TTG AAA ATC ACA TCT GGT GGA GTA					
A K L N D T L L M C L K I T S G G V					
765	774	783	792	801	810
ATT TTC CGG TCA CCT CTA ATG TCA GTT CAG CCC ATA AAT ATG GTG AAG CCT GAT					
I F R S P L M S V Q P I N N M V K P D					
819	828	837	846	855	864
CCA CCA TTA GGT TTG CAT ATG GAA ATC ACA GAT GAT GGT AAT TTA AAG ATT TCT					
P P L G L H M E I T D D G N L K I S					
873	882	891	900	909	918
TGG TCC AGC CCA CCA TTG GTA CCA TTT CCA CTT CAA TAT CAA GTG AAA TAT TCA					
W S S P P L V P F P L Q Y Q V K Y S					
927	936	945	954	963	972
GAG AAT TCT ACA ACA GAT ATC AGA GAA GCT GAC AAG ATT GTC TCA GCT ACA TCC					
E N S T T V I R E A D K I V S A T S					
981	990	999	1008	1017	1026
CTG CTA GTA GAC AGT ATA CTT CCT GGG TCT TCG TAT GAG GTT CAG GTG AGG GGC					
L L V D S I L P G S S Y E V Q V R G					
1035	1044	1053	1062	1071	1080
AAG AGA CTG GAT GGC CCA GGA ATC TGG AGT GAC TGG AGT ACT CCT CCT GTC TTT					
K R L D G P G I W S D W S T P R V F					
1089	1098	1107	1116	1125	1134
ACC ACA CAA GAT GTC ATA TAC TTT CCA CCT AAA ATT CTG ACA AGT GTT GGG TCT					
T T Q D V I Y F P P K I L T S V G S					
1143	1152	1161	1170	1179	1188
AAT GTT TCT TTT CAC TGC ATC TAT AAG AAG GAA AAC AAG ATT GTT CCC TCA AAA					
N V S F H C I Y K K E N K I V P S K					
1197	1206	1215	1224	1233	1242
GAG ATT GTT TGG TGG ATG ATT TTA GCT GAG AAA ATT CCT CAA AGC CAG TAT GAT					
E I V W W H N L A E K I P Q S Q Y D					
1251	1260	1269	1278	1287	1296
GTT GTG AGT GAT CAT GTT ACC AAA GTT ACT TTT TTC ATT CTG AAT GAA ACC AAA					
V V S D H V S K V T F F N L N E T K					
1305	1314	1323	1332	1341	1350
CCT CGA GGA AAG TTT ACC TAT GAT GCA GTG TAC TGC TGC AAT GAA CAT GAA TGC					
P R G K F T Y D A V Y C C N E H E C					

Figure 1B

1359            1368            1377            1386            1395            1404  
 CAT CAT CGC TAT GCT GAA TTA TAT GTG ATT GAT GTC AAT ATC AAT ATC TCA TGT  
 H H R Y A E L Y V I D V N I N I S C  
  
 1413            1422            1431            1440            1449            1458  
 GAA ACT GAT GGG TAC TTA ACT AAA ATG ACT TGC AGA TGG TCA ACC AGT ACA ATC  
 E T D G Y L T K M T C R W S T S T I  
  
 1457            1476            1485            1494            1503            1512  
 CAG TCA CTT GCG GAA AGC ACT TTG CAA TTG AGG TAT CAT AGG AGC AGC CTT TAC  
 Q S L A E S T L Q L R Y H R S S L Y  
  
 1521            1530            1539            1548            1557            1566  
 TGT TCT GAT ATT CCA TCT ATT CAT CCC ATA TCT GAG CCC AAA GAT TGC TAT TTG  
 C S D I P S I H P I S E P K D C Y L  
  
 1575            1584            1593            1602            1611            1620  
 CAG AGT GAT GGT TTT TAT GAA TCC ATT TTC CAG CCA ATC TTC CTA TTA TCT GGC  
 Q S D G F Y E C I F Q P I F L L S G  
  
 1629            1638            1647            1656            1665            1674  
 TAC ACA ATG TGG ATT AGG ATC AAT CAC TCT CTA GGT TCA CTT GAC TCT CCA CCA  
 Y T M W I R I N H S L G S L D S P P  
  
 1683            1692            1701            1710            1719            1728  
 ACA TGT GTC CTT CCT GAT TCT GTG GTG AAG CCA CTG CCT CCA TCC AGT GTG AAA  
 T C V L P D S V V K P L P P S S V K  
  
 1737            1746            1755            1764            1773            1782  
 GCA GAA ATT ACT ATA AAC ATT GGA TTA TTG AAA ATA TCT TGG GAA AAG CCA GTC  
 A E I T I N I G L L K I S W E K P V  
  
 1791            1800            1809            1818            1827            1836  
 TTT CCA GAG AAT AAC CTT CAA TTC CAG ATT CCC TAT GGT TTA AGT GGA AAA GAA  
 F P E N N L Q F Q I R Y G L S G K E  
  
 1845            1854            1863            1872            1881            1890  
 GTC CAA TGG AAG ATG TAT GAG GTT TAT GAT GCA AAA TCA AAA TCT GTC AGT CTC  
 V Q W K M Y E V Y D A K S K S V S L  
  
 1899            1908            1917            1926            1935            1944  
 CCA GTT CCA GAC TTG TGT GCA GTC TAT GCT GTT CAG GTG CCC TGT AAG AGG CTA  
 P V P D L C A V Y A V Q V R C K R L  
  
 1953            1962            1971            1980            1989            1998  
 GAT GGA CTG CGA TAT TGG AGT AAT TGG AGC AAT CCA GCC TAC ACA GTT GTC ATG  
 D G L G Y W S N W S N P A Y T V V M  
  
 2007            2016            2025            2034            2043            2052  
 GAT ATA AAA GTT CCT ATG AGA GGA CCT GAA TTT TGG AGA ATA ATT AAT GGA GAT  
 D I K V P M R G P E F W R I I N G D

2061	2070	2079	2088	2097	2106
ACT ATG AAA AAG GAG AAA AAT GTC ACT TTA CTT TGG AAG CCC CTG ATG AAA AAT					
T M K K E K N V T L L W K P L M K N					
2115	2124	2133	2142	2151	2160
GAC TCA TTG TGC AGT GTT CAG AGA TAT GTG ATA AAC CAT CAT ACT TCC TGC AAT					
D S L C S V Q R Y V I N H H T S C N					
2169	2178	2187	2196	2205	2214
GGA ACA TGG TCA GAA GAT GTG GGA AAT CAC ACG AAA TTC ACT TTC CTG TGG ACA					
G T W S E D V G N H T K F T F L W T					
2223	2232	2241	2250	2259	2268
GAG CAA GCA CAT ACT GTT ACG GTT CTG CCC ATC AAT TCA ATT GGT GCT TCT GTT					
E Q A H T V T V L A I N S I G A S V					
2277	2286	2295	2304	2313	2322
GCA AAT TTT AAT TTA ACC TTT TCA TGG CCT ATG AGC AAA GTA AAT ATC GTG CAG					
A N F N L T F S W P M S K V N I V Q					
2331	2340	2349	2358	2367	2376
TCA CTC AGT CCT TAT CCT TTA AAC AGC AGT TGT GTG ATT GTT TCC TGG ATA CTA					
S L S A Y P L N S S C V I V S W I L					
2385	2394	2403	2412	2421	2430
TCA CCC AGT GAT TAC AAG CTA ATG TAT TTT ATT ATT GAG TGG AAA AAT CTT AAT					
S P S D Y K L M Y F I I E W K N L N					
2439	2448	2457	2466	2475	2484
GAA GAT GGT GAA ATA AAA TGG CTT AGA ATC TCT TCA TCT GTT AAG AAG TAT TAT					
E D G E I K W L R I S S S V K K Y Y					
2493	2502	2511	2520	2529	2538
ATC CAT GAT CAT TTT ATC CCC ATT GAG AAG TAC CAG TTC AGT CTT TAC CCA ATA					
I H D H F I P I E K Y Q F S L Y P I					
2547	2556	2565	2574	2583	2592
TTT ATG GAA CGA GTG GGA AAA CCA AAG ATA ATT AAT AGT TTC ACT CAA GAT GAT					
F M E G V G K P K I I N S F T Q D D					
2601	2610	2619	2628	2637	2646
ATT GAA AAA CAC CAG AGT GAT GCA GGT TTA TAT GTA ATT GTG CCA GTA ATT ATT					
I E K H Q S D A G L Y V I V P V I I					
2655	2664	2673	2682	2691	2700
TCC TCT TCC ATC TTA TTG CTT GGA ACG TTA TTA ATA TCA CAC CAA AGA ATG AAA					
S S S I L L G T L L I S H Q R M K					
2709	2718	2727	2736	2745	2754
AAG CTA TTT TGG GAA GAT GTT CCG AAC CCC AAG AAT TGT TCC TGG GCA CAA GGA					
K L F W E D V P N P K N C S W A Q G					

2763            2772            2781            2790            2799            2808  
CTT AAT TTT CAG AAG AAA ATG CCT GGC ACA AAG GAA CTA CTG GGT GGA GGT TGG  
-----  
L N F Q K K M P G T K E I L L G G G G W  
  
2817            2826            2835            2844            2853            2862  
TTG ACT TAG GAA ATG CTT GTG AAG CTA CGT CCT ACC TCG TGC GCA CCT GCT CTC  
-----  
L T \* E M L V K L R P T S C A P A L  
  
2871            2880  
CCT GAG GTG TGC ACA ATG 3'  
-----  
P E V C T M